

## (*E*)-Phenyl- and -heteroaryl-substituted *O*-benzoyl- (or acyl)oximes as lipoprotein-associated phospholipase A<sub>2</sub> inhibitors

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**Abstract**—A series of (*E*)-phenyl- and -heteroaryl-substituted *O*-benzoyl- (or acyl)oximes **3a–n** were synthesized for evaluating their human lipoprotein-associated phospholipase A<sub>2</sub> (Lp-PLA<sub>2</sub>) inhibitory activities. The less lipophilic derivatives **3a–c** showed the most potent in vitro inhibitory activity on human Lp-PLA<sub>2</sub>.

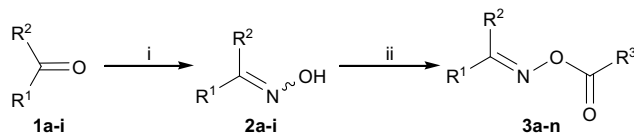
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In general, oxidized low-density lipoproteins (ox-LDLs) play a key role in the early stages of atherosclerosis.<sup>1</sup> Lipoprotein-associated phospholipase A<sub>2</sub> (Lp-PLA<sub>2</sub>) associates predominantly with LDL in plasma and hydrolyzes the *sn*-2 fatty acid of oxidatively modified LDL to generate lysophosphatidylcholine (lyso-PC) and oxidized free fatty acids (oxFFA).<sup>2</sup> In vitro studies have demonstrated that lyso-PC in ox-LDLs is a strong chemoattractant for human monocytes and promotes the chronic inflammation that is associated with macrophage accumulation. Then, macrophages secrete Lp-PLA<sub>2</sub> through the positive feedback mechanism driving progression of atherosclerosis.<sup>3</sup> Therefore, Lp-PLA<sub>2</sub> enzyme is a very attractive target for the treatment of atherosclerosis.

So far, the rational design of Lp-PLA<sub>2</sub> inhibitor is somewhat difficult because the three-dimensional structure of Lp-PLA<sub>2</sub> enzyme has not been elucidated. However, the enzyme is known to be a serine lipase with a catalytic triad that is formed by a histidine and aspartic or glutamic acid at the active site.<sup>4</sup> Thirkettle and co-workers reported that SB-253514 was isolated from *Pseudomonas fluorescens* DMS 11579 and has shown potent inhibitory activity against Lp-PLA<sub>2</sub>.<sup>5</sup> Also, Smith and co-workers

developed a novel series of pyrimidine derivatives through high throughput screening.<sup>6</sup> To explore novel Lp-PLA<sub>2</sub> inhibitors, we screened 4480 compounds that were deposited in Korea Chemical Bank to select (*E*)-benzaldehyde *O*-benzoyloxime (**3a**) with IC<sub>50</sub> value of 3.8 μM. In this letter, we wish to describe the synthesis and in vitro Lp-PLA<sub>2</sub> inhibitory activity of (*E*)-benzaldehyde *O*-benzoyloxime (**3a**) and its derivatives by optimization studies.

A series of (*E*)-phenyl- and -heteroaryl-substituted *O*-benzoyl- (or acyl)oximes **3a–n** were synthesized according to the methods shown in Scheme 1.<sup>7</sup> Treatment of various aldehydes or ketones (**1a–i**) with hydroxylamine hydrochloride and Et<sub>3</sub>N gave the mixture of (*E*)- and (*Z*)-oximes **2a–i** with a high ratio in 60–95% yields, as shown in Table 1. Reaction of the mixture of (*E*)- and (*Z*)-oximes **2a–i** with benzoyl or acyl chlorides gave only an (*E*)-isomer **3a–n** because (*Z*)-isomer could be isomerized to the (*E*)-isomer by triethyl ammonium hydrochloride,<sup>8</sup> as shown in Scheme 1 and Table 2.



**Scheme 1.** Reagents and conditions: (i) NH<sub>2</sub>OH·HCl, Et<sub>3</sub>N, EtOH, rt; (ii) R<sup>3</sup>COCl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C.

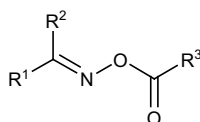
**Keyword:** Lp-PLA<sub>2</sub> inhibitor.

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**Table 1.** Synthesis of (*E*)- or (*Z*)-aldoximes **2a–i**

Compd <b>2</b>	R <sup>1</sup>	R <sup>2</sup>	Yield (%) <sup>a</sup> ( <i>E/Z</i> ) <sup>b</sup>
<b>2a</b>	Ph	H	80 (9/1)
<b>2b</b>	3,4-(F) <sub>2</sub> -Ph	H	95 (9:1)
<b>2c</b>	4-F-Ph	H	95 (8:1)
<b>2d</b>	(3,5-Di- <i>t</i> -Bu-4-OMe)-Ph	H	60 (6:1)
<b>2e</b>	PhCH=CH	H	95 (1.7:1)
<b>2f</b>	Furanyl	H	55 (1.2:1)
<b>2g</b>	Thiophenyl	H	65 (1.2:2)
<b>2h</b>	Ph	Me	80 (9:1)
<b>2i</b>	Ph	Ph	84

<sup>a</sup> Isolated yields from **1a–i**.<sup>b</sup> Isolated ratios.**Table 2.** Lp-PLA<sub>2</sub> inhibitory activities of **3a–n**

Compd <b>3</b>	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	Yield (%) <sup>a</sup>	IC <sub>50</sub> (μM) <sup>b</sup>
<b>3a</b>	Ph	H	Ph	65	3.8
<b>3b</b>	3,4-(F) <sub>2</sub> -Ph	H	Ph	80	2.0
<b>3c</b>	4-F-Ph	H	Ph	80	4.4
<b>3d</b>	(3,5-Di- <i>t</i> -Bu-4-OMe)-Ph	H	Ph	50	11% <sup>c</sup>
<b>3e</b>	PhCH=CH	H	Ph	85	25
<b>3f</b>	Furanyl	H	Ph	72	26.0
<b>3g</b>	Thiophenyl	H	Ph	75	11.2
<b>3h</b>	Ph	Me	Ph	80	29% <sup>c</sup>
<b>3i</b>	Ph	Ph	Ph	77	NA <sup>d</sup>
<b>3j</b>	3,4-(F) <sub>2</sub> -Ph	H	9( <i>Z</i> )-C <sub>17</sub> H <sub>33</sub>	46	16% <sup>c</sup>
<b>3k</b>	3,4-(F) <sub>2</sub> -Ph	H	9( <i>Z</i> ), 12( <i>Z</i> )-C <sub>17</sub> H <sub>31</sub>	24	20% <sup>c</sup>
<b>3l</b>	3,4-(F) <sub>2</sub> -Ph	H	C <sub>9</sub> H <sub>19</sub>	50	12% <sup>c</sup>
<b>3m</b>	3,4-(F) <sub>2</sub> -Ph	H	Ph (4-NO <sub>2</sub> )	65	17% <sup>c</sup>
<b>3n</b>	3,4-(F) <sub>2</sub> -Ph	H	Ph (3,4-F <sub>2</sub> )	85	38

<sup>a</sup> Isolated yields from **2**.<sup>b</sup> Using isolated LDL. Data are shown as mean values of two independent experiments performed in duplicate.<sup>c</sup> Percentage at 25 μM.<sup>d</sup> NA = not active.

The potential of **3a–n** was evaluated as an inhibitor of Lp-PLA<sub>2</sub> (LDL-PLA<sub>2</sub>). Because the plasma isoform of Lp-PLA<sub>2</sub> is 85–90% bound to LDL,<sup>9</sup> the LDL isolated from the plasma of normal lipidemic volunteers<sup>10</sup> was used as the source of enzyme. Then, the amount of [<sup>3</sup>H]acetate produced from [<sup>3</sup>H]PAH (1-*O*-hexadecyl-acetyl-<sup>3</sup>H(N)-phosphatidylcholine) was determined by scintillation counting to reveal the Lp-PLA<sub>2</sub> inhibitory activity.<sup>11</sup> The Lp-PLA<sub>2</sub> inhibitory activities of **3a–n** were confirmed by the positive control with SB381320 supplied by GlaxoSmithKline. Then, SB381320 inhibited Lp-PLA<sub>2</sub> (LDL-PLA<sub>2</sub>) with IC<sub>50</sub> value of 8.8 nM (IC<sub>50</sub> value of 8.0 nM in recombinant Lp-PLA<sub>2</sub> and 67% inhibition in whole human plasma at 100 nM).<sup>12</sup> The data for all compounds **3a–n** has been shown in Table 2. Compound **3a–c** showed an encouraging inhibitory activity against Lp-PLA<sub>2</sub> with IC<sub>50</sub> values of 3.8, 2.0, and 4.4 μM, respectively. The styryl and heterocycle

derivatives **3e–g** at R<sup>1</sup> proved a little less potent than **3a**. On the other hand, a heavily substituted phenyl derivative **3d** was almost devoid of activity. Compounds **3h** and **3i**, which were substituted with methyl and phenyl groups at R<sup>2</sup>, had little effect on potency against Lp-PLA<sub>2</sub>. The highly lipophilic C<sub>9</sub> or C<sub>18</sub> derivatives **3j–l** at R<sup>3</sup> showed a weak inhibitory activity, even though increasing the length of the alkyl chain increased activity.<sup>13</sup> Substitution at R<sup>3</sup> gave compound **3m** with little activity, however, 3,4-difluorophenyl derivative **3n** at R<sup>3</sup> showed a somewhat increased inhibitory activity. These results may be rationalized that Lp-PLA<sub>2</sub> inhibitory activity depends on lipophilicity of the functional groups at R<sup>1</sup> and R<sup>3</sup>.

In conclusion, we have discovered a novel series of Lp-PLA<sub>2</sub> enzyme inhibitors, (*E*)-phenyl- and -heteroaryl-substituted *O*-benzoyl- (or acyl)oximes **3a–n**. Furthermore, the efficacy test of anti-atherogenic activity will be the subject of future publications.

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- Cho, B. R.; Chung, H. S.; Cho, N. S. *J. Org. Chem.* **1998**, *63*, 4685. Typical procedure for the preparation of (*E*)-**3a**: To a solution of benzaldehyde (5.0 mL, 49.0 mmol) in EtOH (50 mL) was added hydroxylamine hydrochloride (4.4 g, 64.0 mmol) and Et<sub>3</sub>N (9.0 mL, 64.0 mmol) at room temperature. After being stirred for 1 h, the reaction mixture was evaporated under reduced pressure to give the residue, to which was added H<sub>2</sub>O (50 mL) and extracted with EtOAc. The combined organic layer was washed with

- brine, dried over anhydrous  $\text{MgSO}_4$ , filtered, and evaporated to give the crude product. Purification by flash column chromatography on silica gel *n*-hexane–EtOAc (1:1) gave the pure (*E*)-**2a** (3.0 g, 50%) and (*Z*)-**2a** (0.32 g, 5%). To a solution of (*E*)-**2a** (0.22 g, 1.8 mmol) in  $\text{CH}_2\text{Cl}_2$  (10 mL) was added benzoyl chloride (0.3 mL, 2.3 mmol) in the presence of  $\text{Et}_3\text{N}$  (0.32 mL, 2.3 mmol) at 0 °C. After being stirred for 1 h, the reaction mixture was quenched with 1 N HCl and extracted with  $\text{CH}_2\text{Cl}_2$  to give the residue, which was washed with saturated  $\text{NaHCO}_3$  aqueous solution and brine and dried over anhydrous  $\text{MgSO}_4$ . Purification by flash column chromatography on silica gel [*n*-hexane–EtOAc (1:1)] gave the pure (*E*)-**3a** (0.26, 65%) as colorless prisms, mp 98–105 °C (*n*-hexane– $\text{CH}_2\text{Cl}_2$ );  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.48 (m, 5H), 7.62 (m, 1H), 7.82 (dd,  $J$  = 1.8, 7.8 Hz, 2H), 8.14 (dd,  $J$  = 2.1, 8.4 Hz, 2H), 8.57 (s, 1H).
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  11. Lp-PLA<sub>2</sub> the enzyme is also known as platelet-activating factor acetylhydrolase (PAF-AH), activity was measured using [ $^3\text{H}$ ] PAF as a substrate.<sup>13,16</sup> Briefly, a micelle substrate was prepared with unlabeled PAF and [ $^3\text{H}$ ] PAF (100  $\mu\text{Ci/mL}$ , 21.5 Ci/mmol, NET 910) in 10 mM phosphate-buffered saline (PBS), pH 7.4, containing 2.7 mM EDTA (PBS–EDTA). The reaction mixture, containing 20  $\mu\text{L}$  of diluted human LDL (4–5  $\mu\text{g}$  protein), 120  $\mu\text{L}$  of PBS–EDTA, and 20  $\mu\text{L}$  of test sample, was preincubated at 37 °C for 15 min. The reaction was initiated by the addition of 40  $\mu\text{L}$  micelle substrate (0.05  $\mu\text{Ci}$ , final concn 80  $\mu\text{M}$  PAF) to measure initial rates of PAF-AH activity. The reaction was stopped by vortexing with 600  $\mu\text{L}$  of  $\text{CHCl}_3/\text{MeOH}$  (2:1) and the  $\text{CHCl}_3$  and aqueous layers were separated by centrifugation. The aqueous layer was removed (250  $\mu\text{L}$ ) and vortexed with 250  $\mu\text{L}$  of  $\text{CHCl}_3$ . The aqueous layer was again removed and the [ $^3\text{H}$ ] acetate determined by scintillation counting (1450 Microbeta Trilux, Qallac Oy, Turku, Finland). The raw counts were corrected for background using a nonenzyme-containing blank and were expressed as nanomoles of PAF degraded per hour per milligram of protein.
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